

Transfection of adipose derived primary MSC using K2 Transfection System

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Materials:

- DMEM, high glucose, GlutaMAX™ Supplement, HEPES (Gibco)
 - Fetal Bovine Serum (Sigma)
 - PBS
 - Trypsin solution
 - Sterile Eppendorf tubes
 - 75T cell culture flask (BD bioscience)
 - 24-well plate (BD bioscience)
 - Plasmid DNA: pE-GFP-C1 (Clontech)
1. Primary human adipose tissue derived Mesenchymal Stem Cells (MSC) were grown in DMEM supplemented with 15% FBS and 1% penicillin-streptomycin in T75 cell culture flask. Cells were trypsinized after 80% confluence and seeded on 24-well plate in density $10^4/\text{cm}^2$ (2×10^4 per well in 0,5 ml DMEM).
 2. Next day K2 multiplier (5 μ l) was added to cells 2h prior transfection. Transfection mixtures were prepared 20 minutes prior transfection according to Table I and following protocol:

Table I.

DNA:K2 ratio- **1:2**

Plate: **24-well plate**

Amount of medium per well: **0,5ml**

K2 multiplier: 10 μ l, 2h incubation

DNA:K2 ratio								
1:2			1:3			1:4		
DNA [μ g]	K2 [μ l]	DMEM [μ l]	DNA [μ g]	K2 [μ l]	DMEM [μ l]	DNA [μ g]	K2 [μ l]	DMEM [μ l]
0,2	0,4	30+30	0,2	0,6	30+30	0,2	0,8	30+30
0,4	0,8	30+30	0,4	1,2	30+30	0,4	1,6	30+30
0,6	1,2	30+30	0,6	1,8	30+30	0,6	2,4	30+30

The indicated amount of K2 transfection reagent (See Table I) was added to 30 μ l of serum-free cell culture medium (DMEM) in a 1.5 ml Eppendorf centrifuge tube. In a separate tube indicated amount of the plasmid DNA (pE-GFP-C1) was mixed with 30 μ l of serum-free medium. The media containing K2 and DNA were combined and mixed by pipetting up and down 5 times. The tubes were incubated at room temperature for 20 min for lipid-DNA complexes to form. At the end of the incubation time, the lipid-DNA complex mixtures (60 μ l per well) were pipetted onto the cells, mixed by tilting the plates a few times and thereafter incubated at 37°C incubator under 5% CO₂. After 24h cells were trypsinised analyzed by flow cytometry.

